A noteworthy Diderma species from Tanzania

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An unidentified species of Diderma, new to Tanzania, is discussed and illustrated. A scanty specimen developed in a moist chamber culture prepared from bark of living Juniperus procera. This moist chamber was cultured for the first time in June 1989 and produced then no Diderma species after one month of incubation. The dried petri dish was stored for ten years in a closed, dark laboratory cupboard and was rewetted in May 1999. After 60 days of incubation an interesting Diderma species developed with walnut-shaped spores bearing an equatorial ring or two perpendicular rings. This specimen may represent a new species, but also seems to have affinity to Diderma punense. We have not been able to obtain the type specimen of the latter species from India to compare it to our specimen. For comparison, some specimens of Diderma cor-rubrum with similar spore morphology were studied with SEM.

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Introduction

In May–June 1989, during the rainy season, the second author and Ms. Tiina Saarimäki collected mushrooms and myxomycetes in Tanzania. In addition to field collections material was collected for moist chamber cultures. The Department of Ecology and Systematics, University of Helsinki made a total of eight field trips in 1988–1995 to different parts of Tanzania. Results of the myxomycete collections made during these trips were published in Härkönen and Saarimäki (1991, 1992, 1994), Ukkola and Härkönen (1996), Ukkola et al. (1996), Ukkola (1998a, b,c), Ukkola (2000), and Härkönen and Ukkola (2000).

In May 1999 the authors rewetted some of the moist chamber cultures that were incubated for the first time in 1989. In one of the cultures an interesting Diderma species developed. We have not been able to identify it so far, but since the species has very interesting spores, we describe, illustrate and discuss this taxon and other closely related Diderma species. For comparison, specimens of Diderma cor-rubrum T. Macbr. were studied with SEM.

Material and methods

Moist chamber cultures were prepared with bark samples collected from living Tanzanian trees in May–June 1989 and wetted in Finland on 6 June 1989. The petri dishes were kept in an incubator at 25°C, and lit artificially in a 12:12 h cycle. During the incubation, mature sporangia were removed. After one month the dishes were dried and stored for ten years in a closed, dark laboratory cupboard. In May 1999 the chambers were rewetted and incubated in normal room conditions in diffuse daylight for two months. To prevent any new contaminations, the wetting and any other opening of the cultures, was made in a sterile room radiated beforehand with UV light. In one of the rewetted cultures after 60 days of incubation a Diderma species developed. The specimen is scanty, consisting of only fifteen mature sporangia. A third rewetting of the culture no further produced material, and the spores did not germinate in a hanging drop of water (see Farr 1981), nor on oat agar. The specimen is deposited at the Botanical Museum of the University of Helsinki (H). It is too scanty to serve as a type specimen of a new species (and its affinity to Diderma punense Patil, Mishra & Ranade (as 'punen-
sis) is still unclear) but, as it is very unique and well matured, we describe it here.

For SEM studies some dried sporangia were dehydrated with 0.1 M phosphate buffer (pH 7.3), and fixed in 2.5% glutaraldehyde at +4°C overnight. After proper rinsing in buffer the sporangia were put in an ethanol series (50% 2 × 5 min, 70% for 24 h, 94% 2 × 10 min, 100% for 24 h), after which they were dried with critical point using Balzers CPD 020 Critical Point dryer. Sporangia were mounted with graphite glue on aluminium stubs and coated with platinum in a Jeol Fine Coat JFC-1100 sputter (25 sec.), and observed with a Jeol JSEM-820 scanning electron microscope using 10 kV in the Institute of Biotechnology, Electron Microscopy Unit, University of Helsinki.

The Tanzanian Diderma species Figs. 1–8


Sporangia in a small group, stalked, hemispherical, depressed and wrinkled below, white to cream-coloured; total height 0.8–1.2 mm, ca. 0.5–0.7 mm in diameter. Stalk thick, about half or little more of the total height, pale staw-yellow, knobbly from protruding calcareous nodes, filled with large, tetragonal and rounded lime crystals. Hypothallus inconspicuous, common to the group. Peridium double, outer layer whitish, egg shell-like, composed of closely aggregated, globose calcareous granules, inner layer thin, whitish on the upper parts, pale yellowish brown and divided with pale bands on the lower parts. Columella small, depressed, pulvinate, round when seen above, concolorous with the stalk. Capillitial threads sparse, thin, pale brown to nearly hyaline, occasionally with a few brown nodules. Spores black in mass, purple-brown by transmitted light, walnut-shaped with an equatorial, saturnoid ring, or often with two rings arranged perpendicularly to each other, very minutely warted or roughened, nearly smooth by light microscope, with SEM the outer layer of the spore wall roughened, “fluffy” often scaling away and exposing the inner, roughened wall; spores 12–14.3–17 μm in diameter.

Discussion

The peeling of the outer spore wall was visible only in SEM (Figs. 5–8), by light microscope with oil-immersion there was no sign of it. In spite of fixing with glutaraldehyde, it is probable that the peeling is an artefact caused by preparatory treatments for SEM.

The Tanzanian Diderma species seems to be closest to Diderma punense, other related species are Diderma cor-rubrum, Diderma marieae.
Figs. 3–8. SEM micrographs of *Diderma* sp. from Tanzania. – 3. Sporangium. Note the knobs on the surface of the stalk caused by large, protruding lime granules. Bar = 100 μm. 4. Globose calcareous granules forming the upper egg-shell-like layer of the peridium. Bar = 1 μm. 5. One spore with roughened, fluffy outer surface, and an equatorial, saturn-like ring. Note the less distinct other ring at right angles to the first. Bar = 1μm. 6. One spore with two clear rings. Bar = 1μm. 7. Two spores in which the outer spore wall is broken. Bar = 5 μm. 8. One “peeled” spore with roughened wall. Bar = 1μm.
Patil, Mishra & Ranade (as 'marie'), and Diderma reticulosporum Nann.-Bremek., K. Mukerji & R. Pasricha.

*Diderma punense* has a white, calcareous stalk, thick peridium which dehisces along hexagonal markings; small, nearly dome-shaped to globose, pinkish white columella, dark brown capillitial threads, and walnut like, smooth spores with encircling ring, (12.5–)13–15 \( \mu \)m in diameter (Mishra & Ranade 1979). We have not been able to obtain the type specimen of *D. punense* that originates from Empress Garden Pune, New Delhi, India (collected on 20 June 1975). The outer peridial layer of the Tanzanian *Diderma* is smooth (Fig. 3) with no hexagonal plates. The lower inner side, however, has beige areolae, divided by pale bands.

*Diderma cor-rubrum* has a clavate, greyish purple to deep purplish red columella (Fig. 9), spores are globose to ovate, occasionally apiculate at one or both ends, verrucose (Martin & Alexopoulos 1969). According to Keller (1970) the apiculation represents a manifestation in optical section of a continuous ring that encircles the surface of the spore. With SEM it really is possible to see a ring around the spores (Figs. 10–11).

*Diderma mariaeae* has spherical to subspherical, thick-walled spores, 12.8–16 \( \mu \)m in diameter, ornamented with short and stout spines fused to.

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Figs. 9–11. SEM micrographs of *Diderma cor-rubrum*. 9. Dehiscent sporangium showing the clavate columella and the membranous inner peridium (USA, T.E. Brooks 1972). Bar = 100 \( \mu \)m. 10. Two warted spores with equatorial rings (USA, T.E. Brooks 1972). Bar = 1\( \mu \)m. 11. Spore ornamented with bacula and a conspicuous ring (India, Thind 7145). Bar = 1\( \mu \)m.
form an incomplete reticulation and encircled by ridge or ridges (Mishra & Ranade 1979).

_Diderma reticulosporum_ has subovoid to somewhat lemon-shaped spores with an equatorial, somewhat protruding line, ornamented with a rather small-meshed irregular reticulation, 10–10.5 × 13–14 μm in diameter (Nannenga-Bremekamp et al. 1984).


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References


